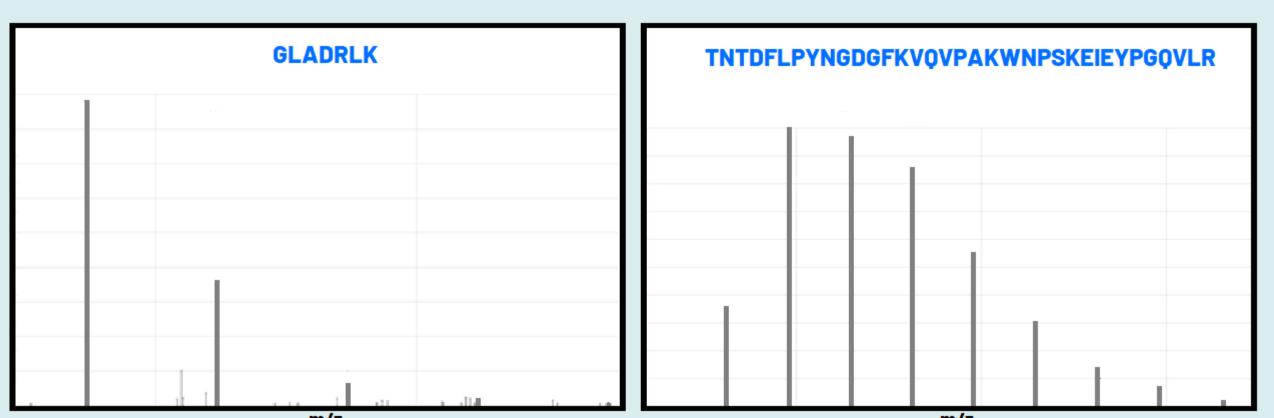
#### How Well Did You Capture that lon? Find Out with PeptidePrisoner! Luis Mendoza, Michael R. Hoopmann, Eric W. Deutsch, and Robert L. Moritz A New Tool in the Trans-Proteomic Pipeline Institute for Systems Biology, Seattle, WA

### Overview

- Establish a quantitative metric for adherence of isotopic envelope of precursor ion to theoretical
- Compute a precursor isolation metric that takes the above fit into account
- User interface for the visualization, exploration, and interrogation of precursor signal quality
- Integrated into the Trans-Proteomic Pipeline (**TPP**) www.tppms.org

## Introduction

- Amino acids in natural or synthetic proteins are composed of atoms with a mixture of **isotopes**.
- This creates an isotopic **envelope** in the MS1 signal of the parent ion.



- A good fit against the theoretically-predicted distribution might provide corroborating evidence for the peptide match.
- A **bad** fit could be indicative of an **incorrect** identification; the wrong charge state reported by the instrument software; **overlapping ions** from other peptides, whether isobaric or not; **noise**; or a combination of these.
- This fit can also be used to estimate the amount of **signal** in the isolation window **attributable** to selected precursor ion.
- A low value is suggestive of co-fragmented species, leading to complex **chimeric** spectra.

## Methods: Precursor sotopic envelope Quality score

- The PINQ score is computed as the best **Chi**squared fit of observed m/z and intensities of isotopic peaks to those **predicted** by theory
- Data are extracted from **pepXML** and **mzML** files

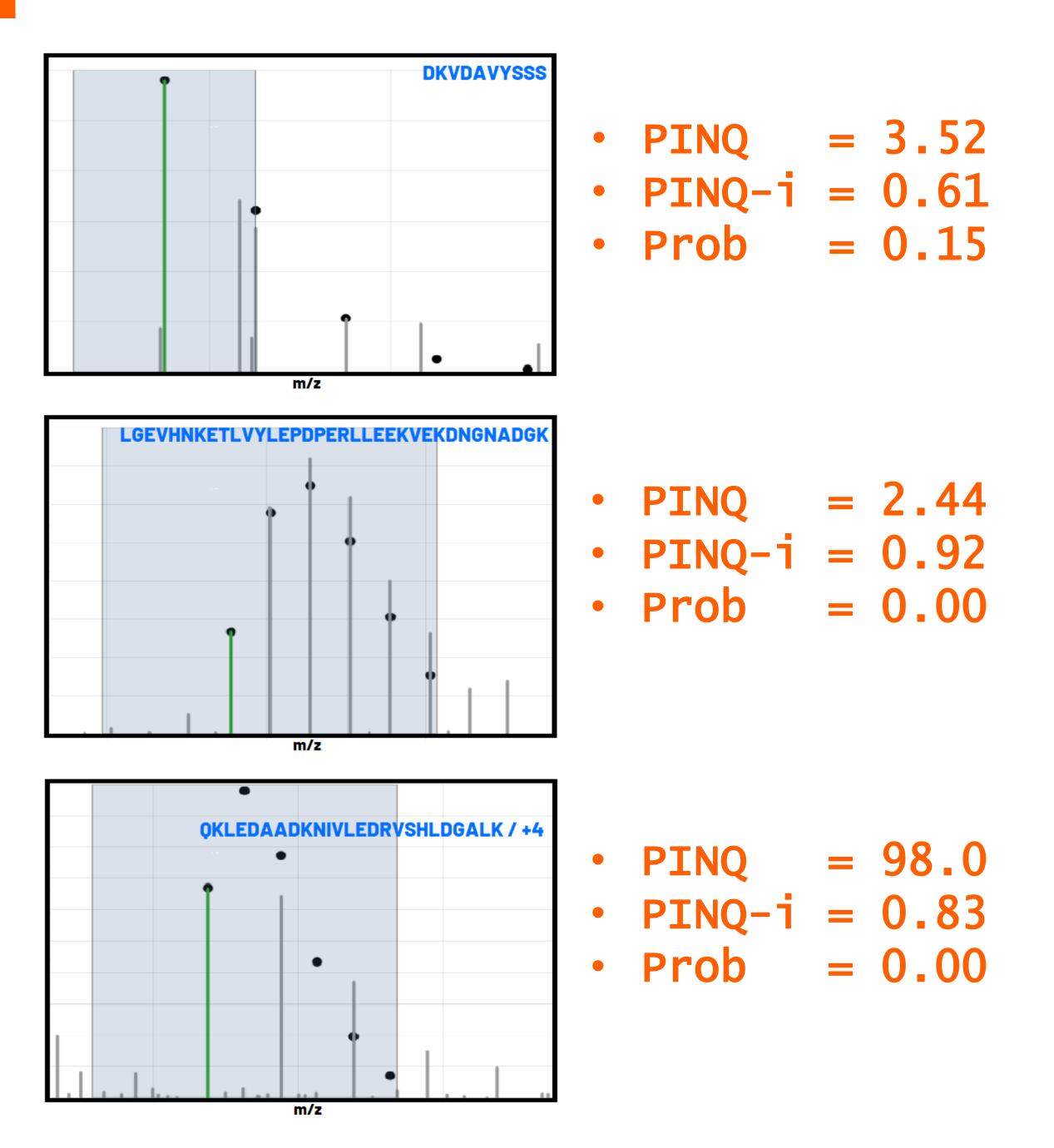
# Methods: PINQ-i (isolation) score

- Computed by adding the peak intensities of all **isotopic peaks** within the selection window, and dividing by the total signal in the window
- The intensity used is the **lower** of:
- observed signal <sup>A</sup>
- intensity attributable via PINQ fit <sup>B</sup>

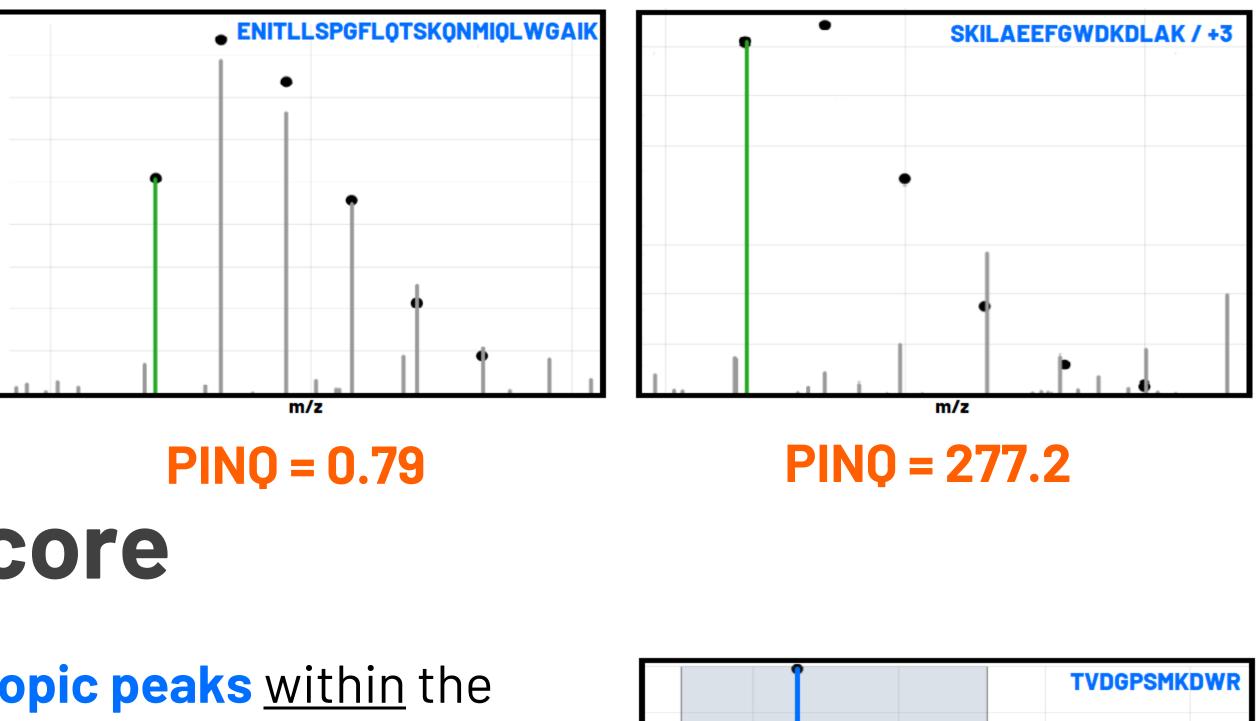
- by the peptide ion

Scores are written back to the source **pepXML** file.

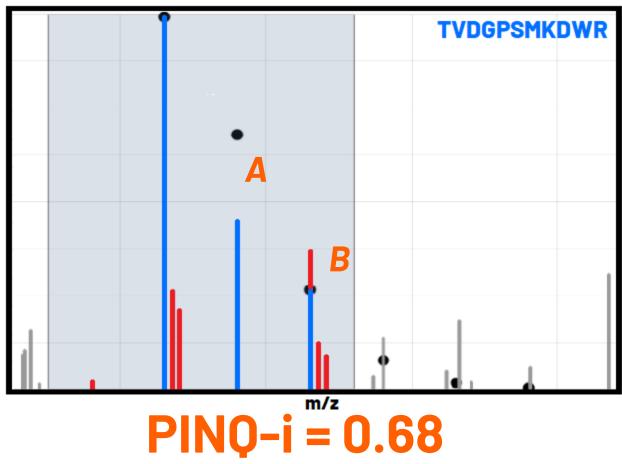
## **Results (examples)**



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**PINQ-i = 0.00** : <u>none</u> of the signal in selection window can be attributed to the identified peptide **PINQ-i = 1.00** : <u>all</u> of the signal was likely generated



Very good isotopic pattern fit, but low probability and marginal isolation score suggest **co**fragmentation of another ion.

Excellent isotopic pattern fit and isolation score. Poor probability suggests this is a real peptide that was **not in the search database** – perhaps a variant, contaminant, or from another organism.

Very poor isotopic pattern fit and probability, and good isolation score would indicate that the charge state might be incorrect. Quick examination via the interface suggests a charge of +2 instead of +4.





#### **User Interface**

**Pr**ecursor **iso**topic envelope explorer

- Peptide and scan information, signal strength
- Relative intensities of detected isotopic peaks as well as those expected from theory
- *PINQ/-i* scores and other TPP confidence metrics
- Links to display the original spectral data along with the expected intensities and best fit
- Pan, zoom, search and filter functionality



## **Future Work**

- Incorporate the PINQ and PINQ-i scores as extra discriminants for PSM validation in **PeptideProphet**
- Flag potentially **chimeric** spectra
- Consider **fragment** ion isotopes

## **Conclusions and Availability**

- The TPP is a widely used and well-validated **free** and open source suite of software tools that facilitates and standardizes proteomics analysis.
- These various updates will allow TPP users to analyze, validate, and visualize precursor isolation quality results from **any** supported search engine.
- These features will be available as of the next release of TPP, version **6.4.0**, planned for Summer 2023, or in preview mode in an upcoming dev build.